# **Tryptone Water**

## **Intended Use**

Tryptone Water is recommended for use in the detection of *Escherichia coli* in food and water samples based on indole production.

## **Summary and Explanation**

Tryptone Water is based on the Tryptone Water formula described in ISO standard 7251.<sup>1</sup> In this procedure, Tryptone Water is used with Lauryl Tryptose (or Sulfate) Broth and EC Broth to determine the most probable number (MPN) of *E. coli* present in the sample. Gas production in both media and indole production in Tryptone Water is used as the basis for this presumptive *E. coli* test.

Tryptone Water may also be used for differentiation of other bacteria based on indole production.

## **Principles of the Procedure**

Tryptone Water contains both tryptone (1%) and sodium chloride. Due to its high tryptophan content, tryptone is suitable for use in detecting indole production by bacteria. Tryptophan is

# **User Quality Control**

Identity Specifications					
Difco <sup>™</sup> Tryptone Water					
Dehydrated Appearance:	White to light beige, free flowing, homogeneous.				
Solution:	1.5% solution, soluble in purified water. Solution is pale to medium amber, clear to slightly opalescent.				
Prepared Appearance:	Light to medium amber, clear to slightly opalescent.				
Reaction of 1.5% Solution at 25°C:	pH 7.3 ± 0.2				

#### Cultural Response Difco™ Tryptone Water

Prepare the medium per label directions. Inoculate and incubate at  $35 \pm 2^{\circ}$ C for 18-24 hours. Add 0.5 mL Indole Reagent (Kovacs) to the tubes to test for indole production. Formation of a red color denotes a positive indole test.

ORGANISM	ATCC™	INOCULUM CFU	RECOVERY	INDOLE PRODUCTION
Enterobacter cloacae	13047	10 <sup>2</sup> -3×10 <sup>2</sup>	Good	-
Escherichia coli	25922	10 <sup>2</sup> -3×10 <sup>2</sup>	Good	+

hydrolyzed and deaminated to produce indole, pyruvic acid and ammonia.<sup>2</sup> Indole can then be detected by the addition of either Kovacs' or Ehrlich's Reagent, which contain an aldehyde group. The aldehyde group combines with indole to produce a red color in the alcohol layer. Sodium chloride is added to the medium to provide a suitable osmotic environment.

## Formula

#### Difco<sup>™</sup> Tryptone Water

Approximate Formula* Per Liter	
Tryptone	g
Sodium Chloride	g
*Adjusted and/or supplemented as required to meet performance criteria.	

## Directions for Preparation from Dehydrated Product

- 1. Dissolve 15 g of the powder in 1 L of purified water.
- 2. Autoclave at 121°C for 15 minutes.
- 3. Test samples of the finished product for performance using stable, typical control cultures.

## **Procedure**

#### Test For Enumeration of Presumptive E. coli

- 1. Suspend one part sample in 9 parts diluent. Homogenize sample.
- 2. Prepare serial 10-fold dilutions to 10<sup>-6</sup> using 1 mL of homogenate and 9 mL of diluent. Mix each dilution thoroughly.
- 3. Transfer 10 mL of test sample or initial suspension to each of 3 tubes of double-strength Lauryl Tryptose Broth (LTB). Repeat using 3 tubes of single-strength LTB. Mix well.
- 4. For each of the serial 10-fold dilutions, transfer 10 mL of test sample to each of 3 tubes of double-strength LTB. Repeat using 3 tubes of single-strength LTB. Mix well.
- 5. Incubate all tubes of LTB at 35-37°C for 24 ± 2 hours and up to 48 hours, if necessary, observing tubes for gas formation.
- 6. Inoculate one 3-mm loopful of broth from each tube in Step 5 showing gas formation to 10 mL of EC Broth warmed to 45°C.
- Incubate the EC Broth tubes in a water bath at 45°C for 24 ± 2 hours (up to 48 hours if necessary), observing for gas formation.
- 8. Inoculate one 3-mm loopful of broth from each tube in Step 7 showing gas formation to 5-10 mL of Tryptone Water warmed to 45°C.



- 9. Incubate Tryptone Water tubes in a water bath at 45°C for 48 hours.
- 10.Add 0.5 mL of Indole Reagent to Tryptone Water tubes, mix well and examine after 1 minute.

#### Indole Determination Using Pure Cultures

- 1. Inoculate Tryptone Water using a light inoculum of an 18-24 hour pure culture.
- 2. Incubate the tubes at  $35 \pm 2^{\circ}$ C with loosened caps for 18-24 hours.
- 3. Add 0.5 mL of Indole Reagent (Kovacs) directly to the tube and agitate. Allow tubes to stand for 5-10 minutes.

## **Expected Results**

#### Test For Enumeration of Presumptive E. coli

For each dilution, record tubes as positive if a red ring forms at the top of the medium indicating indole production. Determine the MPN (Most Probable Number) of *E. coli* present in the sample based on the number of tubes that are positive for both gas and indole. Consult the appropriate 3-tube MPN table.<sup>1</sup>

#### Indole Determination Using Pure Cultures

Examine tubes for the formation of a red ring at the top of the tube indicating indole production.

## **Limitations of the Procedure**

- 1. Detection of *E. coli* in meats using Tryptone Water is a presumptive test. If confirmatory testing is required, please consult appropriate references.
- 2. Indole testing is recommended as an aid in the differentiation of microorganisms based on indole production. For complete identification of the organism, further biochemical evaluation is necessary.

### References

- International Organization for Standardization. 1993. Microbiology general guidance for enumeration of presumptive *E. coli* – most probable number technique. ISO 7251, 1993-12-15, 2nd ed. ISO, Geneva, Switzerland.
- MacFaddin. 2000. Biochemical tests for identification of medical bacteria, 3rd ed. Lippincott Williams & Wilkins, Baltimore, Md.

# Availability

## Difco<sup>™</sup> Tryptone Water

ISO Cat. No. 264410 Dehydrated – 500 g

Difco<sup>™</sup>/BBL<sup>™</sup> Indole Reagent

Cat. No. 261185 Droppers – 50 × 0.5 mL

